

## Report

# A *Drosophila* Gustatory Receptor Essential for Aversive Taste and Inhibiting Male-to-Male Courtship

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## Summary

Contact chemosensation is required for several behaviors that promote insect survival. These include evasive behaviors such as suppression of feeding on repellent compounds, known as antifeedants, and inhibition of male-to-male courtship. However, the gustatory receptors (GRs) required for responding to nonvolatile avoidance chemicals are largely unknown. Exceptions include *Drosophila* GR66a and GR93a, which are required to prevent ingestion of caffeine [1, 2], and GR32a, which is necessary for inhibiting male-to-male courtship [3]. However, GR32a is dispensable for normal taste. Thus, distinct GRs may function in sensing avoidance pheromones and antifeedants. Here, we describe the requirements for GR33a, which is expressed widely in gustatory receptor neurons (GRNs) that respond to aversive chemicals. *Gr33a* mutant flies were impaired in avoiding all nonvolatile repellents tested, ranging from quinine to denatonium, lobeline, and caffeine. *Gr33a* mutant males also displayed increased male-to-male courtship, implying that it functioned in the detection of a repulsive male pheromone. In contrast to the broadly required olfactory receptor (OR) OR83b, which is essential for trafficking other ORs [4], GR66a and GR93a are localized normally in *Gr33a* mutant GRNs. Thus, rather than regulating GR trafficking, GR33a may be a coreceptor required for sensing all nonvolatile repulsive chemicals, including tastants and pheromones.

## Results and Discussion

### Candidate Gustatory Receptors that Function in Contact Chemosensation Identified by DNA Microarrays

The *Drosophila* gustatory receptor (GR) family [5] is distinct from other groups of chemosensory receptors in that it includes members that function in both contact [1–3, 6–11] and noncontact chemosensation [12, 13]. To identify additional GRs that might function in contact chemosensation, which includes taste and the detection of nonvolatile pheromones, we performed DNA microarrays. To conduct this analysis, we took advantage of a mutant, *poxn* [14], in which the chemosensory sensilla (bristles) were transformed to mechanosensory bristles. The major taste organ in the fly is the labellum. Therefore, we prepared RNA from labella dissected from wild-type and *poxn* flies, because enrichment in wild-type labella would therefore indicate that the RNA was

expressed in taste sensilla. The *Gr* RNAs most enriched in the wild-type labella were *Gr66a* and several members of the *Gr64* cluster that are expressed in the sugar-responsive gustatory receptor neurons (GRNs) (see Table S1 available online; enrichments were 6.1- to 14.6-fold). The *Gr* that was the next most enriched in wild-type sensilla was *Gr33a* (Table S1; 5.5-fold enrichment), which was the *Gr* most related to *Gr66a* (Figure S1).

### GR33a Was Expressed in GRNs that Respond to Repulsive Chemicals

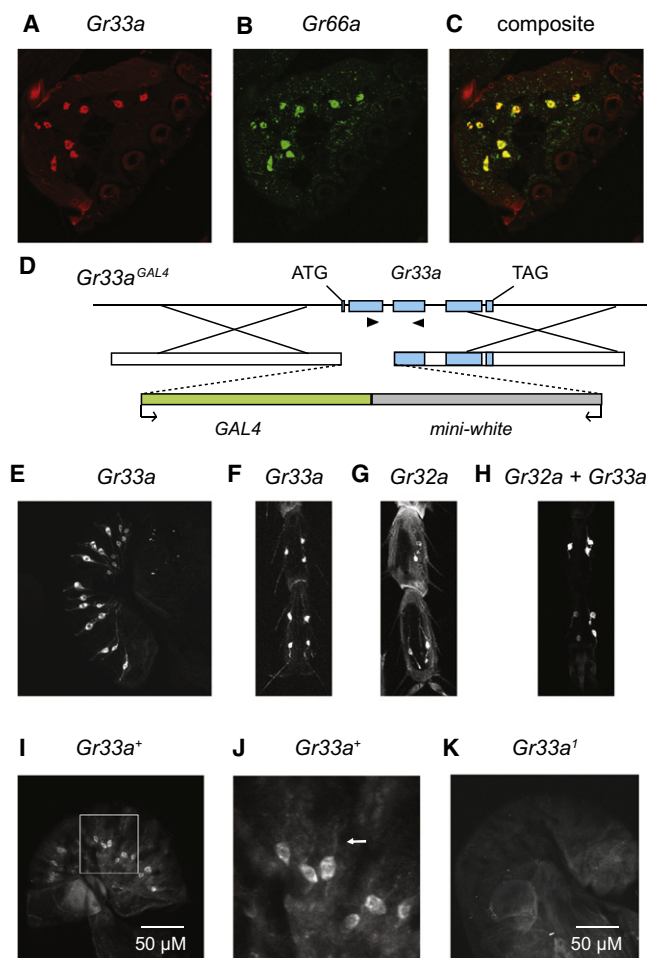
To validate that *Gr33a* was expressed in GRNs, we characterized the spatial distributions of the *Gr33a* RNA, a *Gr33a* reporter, and the endogenous GR33a protein. To examine the *Gr33a* RNA expression pattern, we performed in situ hybridizations and found that ~20 GRNs in the labella expressed *Gr33a* (Figure 1A). Given that GR33a is most related to GR66a and there is a precedent that highly related GRs tend to be expressed in the same GRNs [6, 12, 13], we tested whether the two *Gr* RNAs were coexpressed. We found that all of the *Gr33a*-expressing GRNs expressed the *Gr66a* RNA and vice versa (Figures 1A–1C). Thus, *Gr33a* appeared to be expressed exclusively in GRNs that respond to repulsive chemicals.

To provide a more sensitive reagent to characterize the cellular distribution of *Gr33a*, we generated a *Gr33a* reporter. We used homologous recombination [15] to insert the *GAL4* gene at the site of the normal *Gr33a* translation initiation codon (Figure 1D; *Gr33a*<sup>GAL4</sup>) and assayed for *Gr33a* reporter expression by using the *GAL4/UAS* system [16] in conjunction with a *UAS-mCD8::GFP*. The GFP was expressed in the labellum consistent with the in situ hybridizations (Figure 1E). Examination of multiple confocal sections indicated that the *Gr33a* reporter was expressed in ~20 cells, which was similar to the number of cells positive for the *Gr66a* reporter [17, 18] and *Gr33a* RNA.

Given that leg tarsi also contain gustatory sensilla, we examined the expression of GFP in the legs. The *Gr33a* reporter was expressed in multiple GRNs in the legs. These include seven in the prothoracic legs, four in the mesothoracic legs, and four in the third thoracic legs (Figure 1F). Several of the *Gr33a*-positive GRNs in the prothoracic legs appeared to express the previously described *Gr32a* reporter [3, 18] (Figures 1G and 1H and Table S2). *Gr33a* reporter expression was indistinguishable between males and females (data not shown).

To analyze the subcellular localization of the GR33a protein in GRNs, we raised anti-GR33a antibodies. We found that the anti-GR33a signal was distributed throughout the GR33a-positive GRNs, including the dendrites, axons, and cell bodies (Figures 1I and 1J and Figure S2A). This subcellular distribution of GR33a is reminiscent of GR93a and an odorant receptor (OR), OR83b, both of which are detected in cell bodies and neurites [1, 4]. We did not detect GR33a in the labella of *Gr33a* mutant animals (see below; Figure 1K), indicating that the staining was specific for GR33a. To confirm that the *Gr33a* reporter reflected the endogenous cellular distribution of the GR33a protein, we performed double labeling with

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**Figure 1. Spatial Distribution of *Gr33a***  
(A–C) In situ hybridizations of *Gr33a* and *Gr66a* RNAs. Fluorescent double in situ hybridizations revealed coexpression of *Gr33a* and *Gr66a* RNAs in wild-type labella ( $n = 5$ ). (A) shows *Gr33a* RNA, (B) shows *Gr66a* RNA, and (C) shows a composite of *Gr33a* and *Gr66a* RNAs.  
(D) Structure of *Gr33a* locus and targeting scheme used to generate the *Gr33a*<sup>GAL4</sup> allele by homologous recombination. The arrowheads indicate primers used for the PCR analysis (Figure S3B). The arrows at the bottom indicate the orientations of the *GAL4* and *white* genes.  
(E) *Gr33a* reporter expression in labella ( $n = 10$ ).  
(F–H) Comparison of *Gr33a* and *Gr32a* reporter (*Gr32a*-*GAL4*) expression in male prothoracic legs ( $n = 6$ ). Because we used *GAL4* lines to detect *Gr33a* or *Gr32a* expression, it was not possible to perform double labeling. Therefore, we examined *UAS-mCD8::GFP* expression in flies containing the individual transgenes (*Gr33a*<sup>GAL4/+</sup> or *Gr32a*-*GAL4*) or both together. (F) shows *Gr33a* reporter expression, (G) shows *Gr32a* reporter expression, and (H) shows a combination of both *Gr33a* and *Gr32a* reporter expression.  
(I–K) Expression of the endogenous GR33a protein ( $n = 10$ ). Anti-GR33a immunostaining in *Gr33a*<sup>+</sup> ( $w^{1118}$ ) or *Gr33a*<sup>1</sup> mutant labella is shown. (I) shows the *Gr33a*<sup>+</sup> labellum, and (J) shows an enlarged view of a portion of the *Gr33a*<sup>+</sup> labellum shown in (I) (indicated by the box). The arrow indicates a GR33a-positive dendrite. (K) shows the *Gr33a*<sup>1</sup> allele.

anti-GFP and anti-GR33a antibodies. We found that the same cells were labeled with both antibodies, demonstrating that the *Gr33a* reporter faithfully represented the GR33a expression pattern (Figure S2B). We did not detect a GR33a signal in tissues other than taste organs, including the central brain, fat bodies, gut, testis, and accessory glands with either the *Gr33a* reporter or the anti-GR33a antibodies (data not shown).

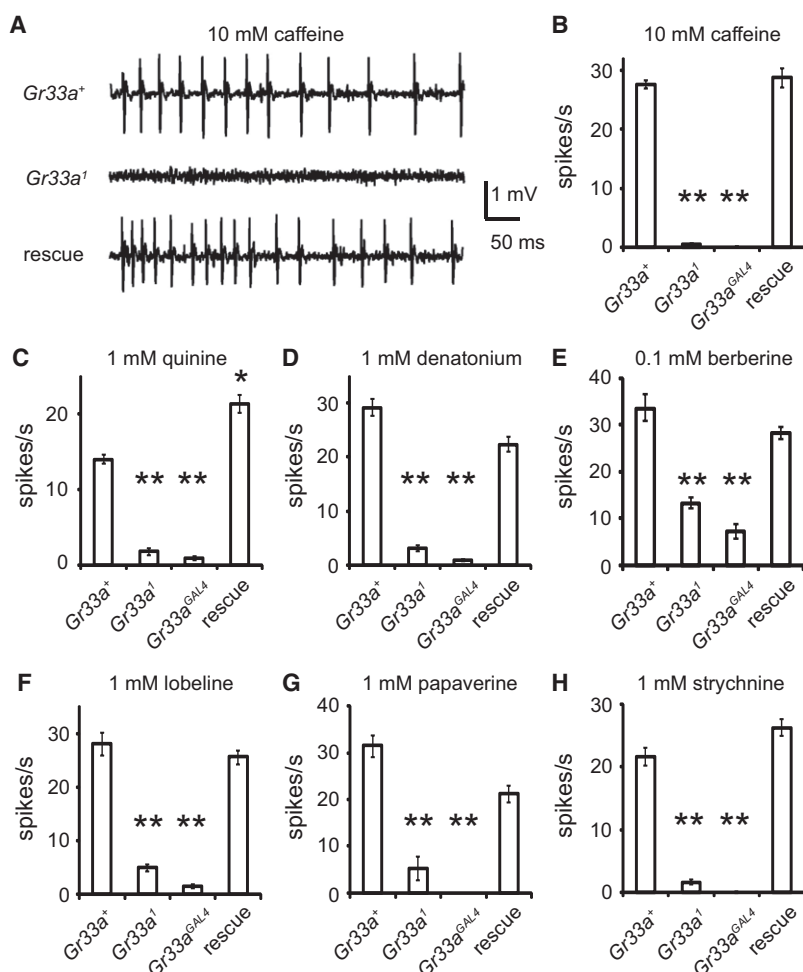
### Broad Requirement for *Gr33a* for the Gustatory Responses to Repellent Chemicals

To characterize the role of *Gr33a*, we created two mutant alleles. The first allele was *Gr33a*<sup>GAL4</sup> because the construct used to obtain the *GAL4* reporter also deleted a portion of the *Gr33a* coding region (Figure 1D; residues 4 to 164). We generated a second allele by homologous recombination, *Gr33a*<sup>1</sup>, which deleted residues 1 to 199 of GR33a (Figure S3A). On the basis of PCR analyses of genomic DNA, the *Gr33a* gene was disrupted as intended in *Gr33a*<sup>1</sup> and *Gr33a*<sup>GAL4</sup> flies (Figure S3B). *Gr66a* RNA was still produced in the mutant GRNs (Figure S3C), indicating that the *Gr33a* mutations did not appear to eliminate the *Gr33a*-expressing GRNs. Both *Gr33a* mutant alleles were homozygous viable (data not shown).

To address whether *Gr33a* null flies have a defect in taste sensation, we performed electrophysiological and behavioral assays. We tested directly for defects in the detection of gustatory stimuli in GRNs by inserting recording electrodes over the tips of taste sensilla and assaying for the production of tastant-induced action potentials. We also tested for behavioral consequences resulting from the *Gr33a* mutations by using two-way choice assays. To perform this latter assay, we placed starved flies in 72-well microtiter dishes with two types of tastants mixed with either red or blue food coloring. Their abdomens appeared red, blue, or purple, depending on whether or not they had a preference for a given tastant. A complete preference for one or the other tastant is indicated by a preference index of 1.0 or 0, and a value of 0.5 results if there is a lack of preference. As expected, given that *Gr33a* was expressed in aversive GRNs, neither *Gr33a*<sup>1</sup> nor *Gr33a*<sup>GAL4</sup> flies showed a significant difference from control flies ( $w^{1118}$ ) in their electrophysiological or behavioral responses to sugars, such as sucrose (Figure S4).

In contrast to the normal sugar responses, *Gr33a* mutant animals showed impairments in the response to all noxious chemical tested. The defects were most pronounced in the tip recording assays. The *Gr33a*<sup>+</sup> control animals ( $w^{1118}$ ) produced action potentials upon presentation of aversive chemicals ranging from caffeine to quinine, denatonium, berberine, lobeline, papaverine, and strychnine (Figure 2 and Figure S5). In contrast, the frequencies of action potentials induced by these tastants were greatly reduced or eliminated in *Gr33a*<sup>GAL4</sup> or *Gr33a*<sup>1</sup> (Figure 2 and Figure S5) or in the transheterozygous flies (*Gr33a*<sup>1</sup>/*Gr33a*<sup>GAL4</sup>; data not shown). The phenotypes were suppressed by introduction of a wild-type transgene (*UAS-Gr33a*<sup>+</sup>) in *Gr33a*<sup>1</sup>/*Gr33a*<sup>GAL4</sup> flies (rescue; Figure 2 and Figure S5), demonstrating that the impairments were due to the mutations in *Gr33a*. Introduction of the rescue construct in the mutant flies caused hypersensitivity to quinine and to the lower concentrations of denatonium, strychnine, and caffeine (Figure S5), possibly as a result of slightly higher expression of GR33a in the transgenic flies than in  $w^{1118}$ . These data support the conclusion that GR33a is required broadly for the avoiding noxious compounds.

To assay the behavioral consequences resulting from disruption of *Gr33a*, we performed two-way choice assays. We performed the assay by comparing the preferences for a lower concentration of a sugar (1 mM sucrose) versus a higher concentration of sugar (5 mM sucrose) in combination with a repulsive tastant, as previously described [19]. Because 5 mM sucrose is preferred over 1 mM sucrose (Figure S4C), a selection of 1 mM sucrose in this assay indicates an aversion to the compound mixed with the 5 mM sucrose.



**Figure 2. Gr33a Was Essential for Production of Action Potentials in Response to Aversive Compounds.**

Shown are the results of tip recordings with s6 sensilla. (A) Representative traces of caffeine-induced nerve firings between 50 and 550 ms after application of the recording electrode. The flies used to test for rescue of the *Gr33a* mutant phenotype by the wild-type *Gr33a<sup>+</sup>* transgene were *Gr33a<sup>1</sup>/Gr33a<sup>GAL4</sup>;UAS-Gr33a*. The *Gr33a<sup>GAL4</sup>* mutant allele did not show a response to caffeine (data not shown).

(B–H) Quantification of the mean action potentials (spikes per second) induced by the indicated noxious chemicals. The genotypes are listed below. The means were based on data collected between 50 and 1050 ms after presentation of the compounds.

The error bars represent standard errors of the mean (SEMs). The data were analyzed with one-way ANOVA followed by Scheffe's post-hoc analysis. The asterisks indicate statistically significant differences from the *Gr33a<sup>+</sup>* control (\**p* < 0.05, \*\**p* < 0.01). Detailed statistics are presented in Table S3. See Figure S5 for dose-response analyses, which include the data summarized in this figure.

### Mutation of *Gr33a* Increased Male-to-Male Courtship

In view of our findings that *Gr33a* mutant flies are unresponsive to all aversive tastants tested, we wondered whether it was also required for inhibiting male-to-male courtship, given that this behavior is suppressed by an inhibitory pheromone present on the male cuticle [20]. This possibility was further supported by the observation that the *Gr32a<sup>+</sup>* gene product, which suppressed male-to-male courtship [3], appeared to be coexpressed in the legs with *Gr33a* (Figures 1F–1H). In support of this conclusion, we detected the same number of GFP-positive (*UAS-*

*mCD8::GFP*) neurons in flies harboring the *Gr33a-GAL4* alone (*Gr33a<sup>GAL4</sup>/+*) or both the *Gr33a<sup>GAL4</sup>/+* and *Gr32a-GAL4* drivers (*Gr33a<sup>GAL4</sup>/+* and *Gr32a-GAL4*; Table S2).

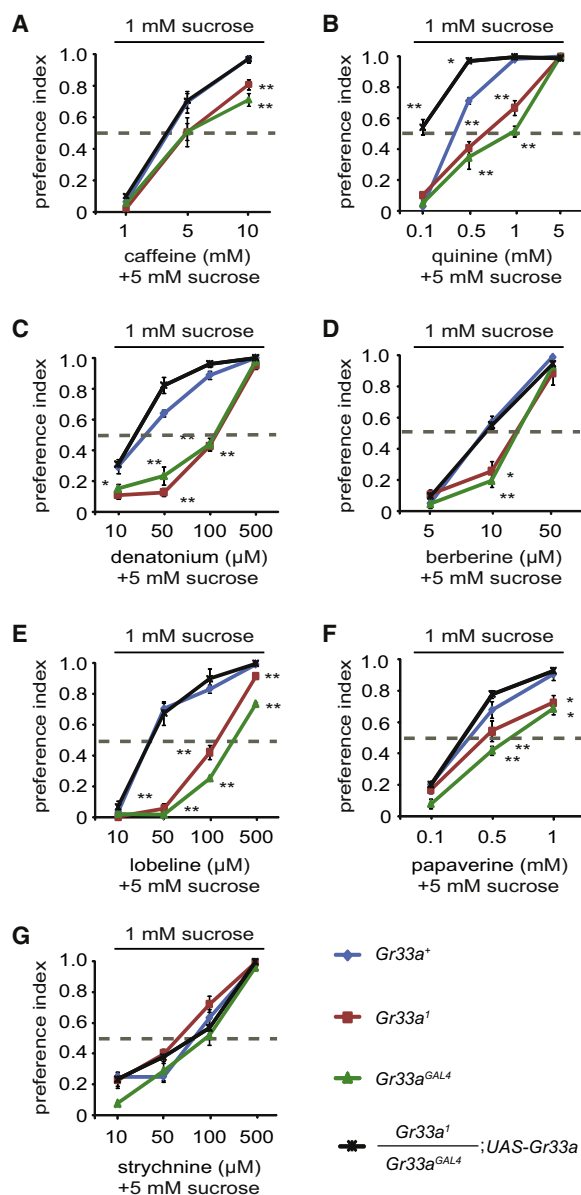
To test whether the *Gr33a* mutation increased male-to-male courtship, we combined *Gr33a<sup>+</sup>* and mutant males with passive, decapitated wild-type males as described [3]. The courtship index was the percentage of time during a 10 min interval that the male engaged in courtship behaviors, including vibration of the wing, licking, and attempted copulation. We found that the *Gr33a<sup>1</sup>* male flies displayed increased courtship toward passive, decapitated males (Figure 4A). This phenotype was rescued by the *Gr33a<sup>+</sup>* transgene (Figure 4A). In contrast, courtship of *Gr33a<sup>1</sup>* males to decapitated or normal females was not different significantly from the *Gr33a<sup>+</sup>* males (Figures 4B–4D). *Gr33a* mutant and *Gr33a<sup>+</sup>* females did not display significant differences in mating (Figures 4E and data not shown). These results raise the possibility that *Gr33a* is required for sensing an inhibitory male pheromone.

### Normal GR Trafficking in the Absence of GR33a

The observation that GR33a is required for responding to every avoidance compound tested raises the possibility that it is corequired with other GRs for the responses to aversive chemicals. Consistent with this proposal, misexpression of *Gr33a* alone in tissue-culture cells or in *Gr5a*-expressing

With one exception (strychnine), we found that the behavioral responses to the repulsive compounds were reduced in both *Gr33a* alleles, and each of the defects was rescued by introduction of the wild-type *Gr33a<sup>+</sup>* transgene (Figure 3). At the higher concentrations of aversive compounds, *Gr33a* mutant flies tended to select 1 mM sucrose over the 5 mM sucrose/bitter compound cocktails. These results are not necessarily inconsistent with the tip recordings in view of the observation that bitter compounds also suppress the ability of S cells to sense sugars [19]. Thus, when 5 mM sucrose is mixed with high concentrations of bitter compounds, the 1 mM sucrose may be perceived as sweeter than the 5 mM sucrose. The absence of a behavioral phenotype in response to strychnine might indicate that the sugar-responsive S cell in the s6 sensilla is very sensitive to inhibition by strychnine. Alternatively, we cannot exclude the possibility that there exists another strychnine receptor that is expressed in a subset of sensilla that we did not assay in the tip recordings. Expression of the *Gr33a<sup>+</sup>* transgene caused the flies to display increased repulsion to quinine (Figure 3B). This was most pronounced in response to the lowest concentration of quinine tested (0.1 mM). The *Gr33a<sup>+</sup>* control and mutant flies strongly preferred the 5 mM sucrose plus 0.1 mM quinine, whereas the transgenic flies expressing the rescue construct no longer had a bias for this 5 mM sucrose/0.1 mM quinine cocktail.





**Figure 3. *Gr33a* Mutant Flies Display Reduced Avoidance to Most Bitter Chemicals**

(A–G) The two-way choice behavioral assays were performed by giving the flies a choice between 1 mM sucrose alone or 5 mM sucrose in combination with the indicated concentrations of aversive chemicals. A value of 1 indicates a complete preference for 1 mM sucrose, and a value of 0 indicates a complete preference for the 5 mM sucrose mixed with the aversive chemical. A value of 0.5 indicates no preference for the two alternative tastants. The strains used were *Gr33a*<sup>+</sup> (*w*<sup>1118</sup>; blue lines), *Gr33a*<sup>1</sup> (red), *Gr33a*<sup>GAL4</sup> (green), and *Gr33a*<sup>1</sup>/*Gr33a*<sup>GAL4</sup>;UAS-*Gr33a* (black). We used one-way ANOVA followed by Scheffé's post-hoc analysis to perform tests for statistical significance. The error bars represent SEMs. The asterisk indicates statistically differences from the *Gr33a*<sup>+</sup> control (\**p* < 0.05). Detailed statistics are presented in Table S4.

GRNs was insufficient to confer sensitivity to any aversive compound tested (S.J.M. and C.M., unpublished data). Thus, the receptors for repellent chemicals may comprise at least two subunits. A minimum of three *Gr*s are required for the response to caffeine: *Gr66a*, *Gr93a* and *Gr33a*; however, mis-expression of all three *Gr*s does not produce a response to

caffeine (S.J.M. and C.M., unpublished data). Thus, the complexity of these receptors may be greater than either the *Drosophila* CO<sub>2</sub> heterodimeric receptor [12, 13] or the mammalian taste receptors, which consist of homo- and heterodimers [21]. Because GR64f functions in concert with GR5a or GR64f for sugar sensation [7], it appears that distinct GRs (GR64f and GR33a) are broadly corequired with other receptors for the responses to sugars and avoidance chemicals.

In the *Drosophila* olfactory system, OR83b is required for trafficking of other ORs in olfactory sensory neurons [4]. To address whether GR33a may play a similar role, we compared the spatial distribution of GR93a in *Gr33a*<sup>+</sup> and *Gr33a* mutant GRNs. As we have shown recently, GR93a is detected in dendrites, axons, and cell bodies of wild-type avoidance GRNs [1] (Figure S6A). We found that anti-GR93a staining was indistinguishable in the *Gr33a* mutant (Figure S6B). To address whether GR66a was dependent on GR33a, we used transgenic flies expressing Myc-tagged GR66a (Myc::GR66a), which rescues the *Gr66a* mutant phenotype [2]. As with GR93a, the Myc-GR66a protein was detected in dendrites, axons, and cell bodies in *Gr33a*<sup>+</sup> GRNs (Figure S6C). This spatial distribution was unchanged in a *Gr33a* mutant background (Figure S6D).

We conclude that GR33a does not function in receptor trafficking, as is the case for OR83b in olfactory receptor neurons. These results raise the possibility that GR33a serves as an obligatory receptor subunit for the detection rather than the trafficking of all repellent compounds that are sensed through contact chemosensation, including aversive tastants and pheromones. Given that three GRs are required but not sufficient for sensing caffeine, the complexity of the aversive GRs may be greater than that of any known chemoreceptor.

## Future Perspective

The identification of GR33a as the first molecular target required broadly for detection of all antifeedants provides the possibility of screening for GR33a inhibitors, which would promote the intake of noxious chemicals and result in improved insect control. Because many GRs such as GR33a are divergent between *Drosophila* and other insects, such as *Anopheles gambiae* (28% identical with GPRGR43) [22, 23], such inhibitors have the potential to selectively target GRs in disease vectors but not in other insects that would be of benefit to maintain.

## Supplemental Data

Supplemental Data include Supplemental Experimental Procedures, five tables, and seven figures and can be found with this article online at [http://www.cell.com/current-biology/supplemental/S0960-9822\(09\)01487-0](http://www.cell.com/current-biology/supplemental/S0960-9822(09)01487-0).

## Acknowledgments

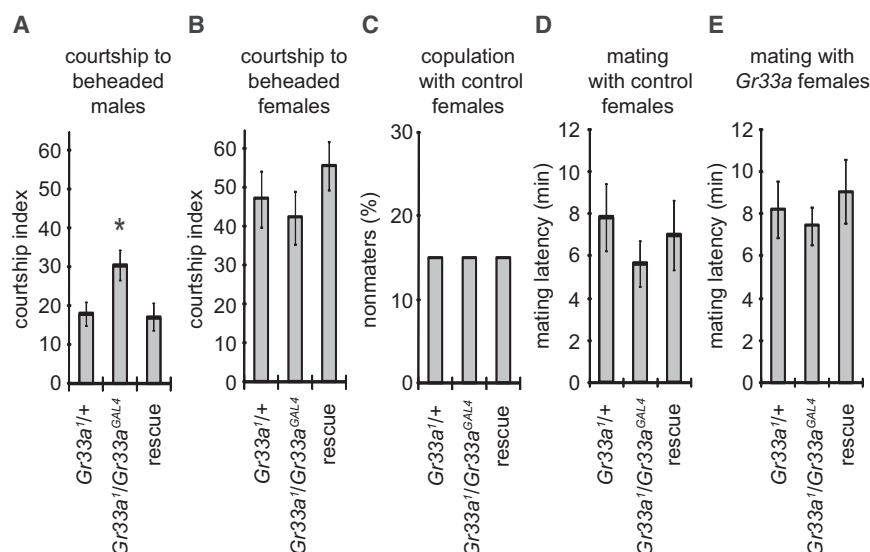
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**Figure 4. Increased Male-to-Male Courtship Behavior in *Gr33a* Mutant Flies**

(A) Courtship behavior between males of the indicated genotypes and passive (beheaded) males. The courtship index is the percentage of time that the intact males exhibited courtship behavior during the 10 min observation period. The error bar represents the SEM.

(B) Courtship behavior exhibited by males of the indicated genotypes to beheaded females.

(C) Fraction of males that did not mate to *Gr33a*<sup>+</sup> intact females.

(D) Time to copulation with *Gr33a*<sup>+</sup> control females.

(E) Average latency time for the *Gr33a*<sup>+</sup> males to mate with indicated females.

ANOVA with the Scheffe's post-hoc tests were used to compare multiple sets of data (A, B, D, and E), and the Chi-square test was used to test the statistical significance of nonmating (C). *p* values < 0.05 were considered statistically significant. The asterisks indicate statistically differences from the *Gr33a*<sup>+</sup> control (\**p* < 0.05, \*\**p* < 0.01). Detailed statistics are presented in Table S5.

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